Linezolid plasma and intrapulmonary concentrations in critically ill obese patients with ventilator-associated pneumonia: intermittent vs continuous administration

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Take-home message: The possibility of underdosing linezolid in obese critically ill patients is high, and continuous infusion may be a useful tool to reduce this risk.

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Abstract Purpose: Clinical application of an antibiotic’s pharmacokinetic/pharmacodynamic (PK/PD) properties may improve the outcome of severe infections. No data are available on the use of linezolid (LNZ) continuous infusion in critically ill obese patients affected by ventilator-associated pneumonia (VAP). Methods: We conducted a prospective randomized controlled trial to compare LNZ concentrations in plasma and epithelial lining fluid (ELF), when administered by intermittent and continuous infusion (II, CI), in obese critically ill patients affected by VAP. Results: Twenty-two critically ill obese patients were enrolled. At the steady state, in the II group, mean ± SD total and unbound maximum–minimum concentrations \( \frac{C_{\text{max}}}{C_{\text{max},u}} - \frac{C_{\text{min}}}{C_{\text{min},u}} \) were \( 6.2 \pm 2.3 \) and \( 4.3 \pm 1.6 \) mg/L, respectively. Within a minimum inhibitory concentration (MIC) range of 1–4 mg/L, the median (IQR) time LNZ plasma concentration persisted above MIC (% \( T > \text{MIC} \) ) was significantly higher in the CI than the II group \( [100 (100–100) \) vs \( 100 (89–100), p = 0.05; 100 (100–100) \) vs \( 82 (54.8–98.8), p = 0.009; 100 (74.2–100) \) vs \( 33 (30.2–78.5), p = 0.005; respectively]. Pulmonary penetration (%) was higher in the CI group, as confirmed by a Monte Carlo simulation \( [98.8 (IQR 93.8–104.3) \) vs \( 87.1 (IQR 78.7–95.4); p < 0.001]. Conclusions: In critically ill obese patients affected by VAP, LNZ CI may overcome the limits of standard administration but these advantages are less evident with difficult to treat pathogens (MIC = 4 mg/L). These data support the usefulness of LNZ continuous infusion, combined with therapeutic drug monitoring (TDM), in selected critically ill populations.

Keywords Linezolid · VAP · Continuous infusion · Pharmacokinetics
Introduction

Ventilator-associated pneumonia (VAP) due to methicillin-resistant Staphylococcus aureus (MRSA) still remains a leading cause of morbidity and mortality. Inadequate antimicrobial treatment against these strains is widely described as a risk factor for worse outcome [1–5].

Linezolid (LNZ) is now considered the first choice for the treatment of MRSA VAP, especially in the presence of strains with vancomycin minimum inhibitory concentration (MIC) values of 1 mg/L or more [6–9]. LNZ acts by a time-dependent antimicrobial killing mechanism: time with plasma concentrations higher than MIC ($T >$ MIC) exceeding 85 % and area under the serum–time concentration curve/MIC (AUC/MIC) more than 80 h are the pharmacodynamic (PD) parameters that best predict the clinical efficacy [10, 11]. LNZ optimally penetrates different organs; however, in critically ill patients plasma and pulmonary concentrations may significantly differ from healthy volunteers [9, 12, 13]. A wide array of pathophysiological changes occurring in critically ill patients may influence antibiotics' pharmacokinetic (PK) properties according to either their lipophilic or hydrophilic nature [14, 15]. Continuous infusion has been proposed as a strategy to minimize the risk of time-dependent antibiotic underexposure in the presence of difficult to treat infections [16–18].

Obesity is now becoming a worldwide healthcare issue and the incidence of obese patients admitted to the ICU has also been increasing. In these patients, many physiological changes may influence antibiotics’ tissue distributions but few data are available to adapt drug dosages and the administration schedule [19, 20]. Data from the literature regarding the LNZ PK in obese critically ill patients are rare and the need to increase LNZ daily dose in accordance with the patients’ mass index (BMI) is now a matter of debate [21–24].

To the best of our knowledge, no information is available on the plasma and pulmonary pharmacokinetics of continuous LNZ infusion use in obese ICU patients with pneumonia. Therefore, we conducted a randomized controlled trial with the aim of comparing the plasma and pulmonary [epithelial lining fluid (ELF)] PK profile of LNZ when administered as intermittent infusion (II) or continuous infusion (CI) in critically ill obese patients with VAP.

Materials and methods

Patients and study design

This study was performed in the 18-bed ICU of a 1,500-bed teaching hospital in Rome, Italy. The protocol was approved by the Catholic University’s Ethical Committee (approval number P/951/CE/2010). Written informed consent was obtained from the patients’ legally authorized representative. Critically ill obese (BMI ≥ 30 kg/m²) adult patients were considered eligible for the study when the attending physician prescribed LNZ as empirical treatment (within 12 h from microbiological pulmonary sampling) of a possible MRSA VAP, in the absence of any exclusion criteria: known LNZ allergy; creatinine clearance less than 40 mL/min (calculated according to the Cocker–Gault formula) apart from those ones who were anuric and on continuous veno-venous hemodiafiltration (CVVHDF); thrombocytopenia (platelet count less than 80,000/mm³); severe hepatic failure (Child–Pugh C); little chance of survival as defined by SAPS II; concomitant treatment with other drugs that can potentially interfere with LNZ (i.e., macrolides, serotonin modulators, omeprazole) [25] [see electronic supplementary material (ESM)]. Patients were randomized (using the opaque sealed envelope method) to receive linezolid (Zyvoxid®; Pfizer, Italia) by intermittent infusion (II) or continuous infusion (CI). The II group received LNZ as a 60-min intermittent intravenous (i.v.) administration (600 mg every 12 h); the CI group received LNZ as 600 mg i.v. loading dose (given in 60 min) followed by 1,200 mg continuous infusion/24 h (50 mg/h). After 2 days of therapy, at steady state, PK analyses of the study group were performed. Thereafter, therapy was continued by standard intermittent dosing. Clinical and demographic data were recorded upon enrollment (see ESM). Safety and adverse events were determined through the observed biochemical abnormalities, documented according to the Department of Health and Human Services–Common Terminology Criteria for Adverse Events (DHHS-CTCAE v.3.0) classification [26].

Sample collection

In the II group blood samples were collected after the fifth dose (on day 3 of treatment) at 70 (immediately before the initiation of the infusion) and 1, 2, 4, 8, 10, and 11 h after the end of the infusion (i.e., 2, 3, 5, 9, 11, and 12 h after the start of infusion). In the CI group blood samples were collected at 48, 53, 57, and 60 h after the first dose (i.e., on day 3 of treatment). According to patients’ respiratory status, one microbronchoalveolar lavage (BAL) (40 mL sterile 0.9 % saline solution was blindly instilled through a telescopic catheter and immediately aspirated in a trap) was performed at steady state.

Pharmacokinetic/pharmacodynamic analysis

Pharmacokinetic parameters were determined by a one-compartment model with first-order elimination. The $0–12 \text{ h } (\text{AUC}_{0–12})$ was determined by the linear trapezoidal rule. $\text{AUC}_{0–24}$ was calculated as $\text{AUC}_{0–12} \times 2$. LNZ...
maximum, minimum, and steady-state concentrations \((C_{\text{max}}, C_{\text{min}}, \text{and } C_{\text{ss}})\) were directly obtained from observed peak, trough, and steady-state concentrations. Epithelial lining fluid (ELF) linezolid \((LNZ_{\text{ELF}})\) concentration was calculated from BAL concentration \((LNZ_{\text{BAL}})\) using urea as dilution marker: \(LNZ_{\text{ELF}} = LNZ_{\text{BAL}} \times \text{urea dilution index (plasma urea concentration/BAL urea concentration)}\) [12]. In all patients receiving II, distribution volume \((V_d)\), drug clearance \((CL)\), and elimination half-life \((t_{1/2})\) were calculated after a single 600-mg intravenous dose at steady state. Time above the minimum inhibitory concentration \((T > \text{MIC})\) of 85 and 100 % and area under the concentration curve \((AUC)_{0-24}/\text{MIC} \) ratio more than 80 h were used as PD targets [11]. Graphing of data was undertaken using Prism version 6.0 for Windows (graphPad Software, San Diego, CA).

Statistical analysis

All statistical analyses were performed using the Intercooled Stata program, version 11 (StatCorp LP). The Kolmogorov–Smirnov test was used to test the variables’ distribution. The data with a non-Normal distribution were assessed with the Mann–Whitney test and the median and selected centiles (25–75th) are given. The data with a normal distribution were assessed with Student’s test. Categorical variables are presented as proportions and were analyzed with the use of the Chi-square test or Fisher’s exact test, as appropriate. A \(p\) value less than 0.05 was considered significant. A power calculation for independent patients with an alpha of 0.05 and a power of 90 %, using a delta (difference of \(C_{\text{min}}\) between population means) of 4 and a sigma (SD) of 200 %, required a sample size of 12 patients. For ELF/plasma ratio results, a Monte Carlo simulation involving 1,000 iterations was also performed [27].

Linezolid assays and microbiological analysis

Plasma and pulmonary LNZ concentrations and microbiological isolates were analyzed as previously reported [28, 29] (see ESM).

Results

Patient demographics

During the study period (April 2011–April 2013) 22 obese critically ill obese patients were enrolled (see Fig. A in the ESM). Eleven patients were randomized to receive LNZ by II and 11 by CI. Patients’ clinical and demographic characteristics are described in Table 1. Median BMI (IQR) was 33.2 kg/m\(^2\) (32.6–37.5) without significant intergroup differences \((p = 0.28)\). The two groups were similar regarding disease severity (SOFa scores), type of admission (55 % medical), and concomitant organ failures (respiratory, cardiovascular, and renal function), but admission SAPS II score was significantly higher in the CI group \((p = 0.02)\). Two patients receiving CI were anuric and underwent CVVHDF during all the infusion period Table 1.

Plasma pharmacokinetic parameters

Total and unbound LNZ plasma concentration versus time curves are shown in Fig. 1 for both groups. At steady

### Table 1 Clinical and demographic data of the 22 enrolled patients

<table>
<thead>
<tr>
<th>Patients’ characteristics</th>
<th>II group ((n = 11))</th>
<th>CI group ((n = 11))</th>
<th>(p) value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years</td>
<td>62.5 ± 10.5</td>
<td>64.7 ± 10.4</td>
<td>0.63</td>
</tr>
<tr>
<td>Male, N (%)</td>
<td>3 (27.3)</td>
<td>5 (45.5)</td>
<td>0.66</td>
</tr>
<tr>
<td>BMI, kg/m(^2) (IQR)</td>
<td>33.3 (32.7–39.1)</td>
<td>33.1 (32.3–34.9)</td>
<td>0.28</td>
</tr>
<tr>
<td>SAPS II</td>
<td>42.7 ± 8.6</td>
<td>54.8 ± 12.5</td>
<td>0.02*</td>
</tr>
<tr>
<td>SOFA</td>
<td>6.4 ± 3.2</td>
<td>5.6 ± 2.8</td>
<td>0.68</td>
</tr>
<tr>
<td>Medical admission, N (%)</td>
<td>7 (63.6)</td>
<td>5 (45.5)</td>
<td>0.7</td>
</tr>
<tr>
<td>(\text{PaO}_2/\text{FiO}_2) (\text{ratio, mmHg (IQR)})</td>
<td>146.1 ± 60</td>
<td>149.2 ± 61</td>
<td>0.9</td>
</tr>
<tr>
<td>Septic shock, N (%)</td>
<td>6 (54.5)</td>
<td>6 (54.5)</td>
<td>1</td>
</tr>
<tr>
<td>Albumin concentration, g/dL</td>
<td>2.6 ± 0.5</td>
<td>2.5 ± 0.6</td>
<td>0.43</td>
</tr>
<tr>
<td>Gram-positive infection</td>
<td>5 (45.5)</td>
<td>2 (18.2)</td>
<td>0.36</td>
</tr>
<tr>
<td>Day 4 clinical improvement, N (%)</td>
<td>8 (72.7)</td>
<td>9 (81.8)</td>
<td>1</td>
</tr>
<tr>
<td>ICU mortality, N (%)</td>
<td>4 (36.4)</td>
<td>1 (9)</td>
<td>0.31</td>
</tr>
</tbody>
</table>

Data are expressed as mean ± SD unless otherwise indicated. II intermittent infusion, CI continuous infusion, IQR interquartile range BMI body mass index, SAPS II simplified acute physiology score II, SOFA sequential organ failure assessment, ICU intensive care unit. *\(p < 0.05\)

a Two patients on continuous renal replacement therapy (CRRT) are not included. Creatinine clearance was calculated according to the Cockcroft–Gault formula.
state, in the II group, mean ± SD $C_{\text{max}}/C_{\text{max,u}}$ and $C_{\text{min}}/C_{\text{min,u}}$ were $10 \pm 3.7/6.8 \pm 2.6$ mg/L and $1.7 \pm 1.1/1.2 \pm 0.8$ mg/L, respectively. In the CI group, the mean ± SD total and unbound plasma concentrations ($C_{ss}$ and $C_{ss,u}$) were $6.2 \pm 2.3$ and $4.3 \pm 1.6$ mg/L, respectively. In the CI group, during all the infusion time, total LNZ plasma concentration was above 4 mg/L and unbound LNZ concentration above 2 mg/L. Otherwise, in the II group, $T_{[MIC]}$ (expressed as dosing intervals percentage) was significantly lower than in the CI group except for the 0.5 mg/L MIC value (total and unbound LNZ) and 4 mg/L MIC value (LNZu), Table 2.

Patients receiving CI, compared to those in the II group, had a significantly higher probability of target attainment (PTA) ($T > 85\%$) at the 2 mg/L MIC value for both total and unbound drug ($100 \text{ vs } 45.5\%, p = 0.02$ and $90.9 \text{ vs } 27.3\%, p = 0.01$, respectively). Similar results were observed for $T > 100\%$ PTA: $100 \text{ vs } 45.5\%, p = 0.02$ (total LNZ and 2 mg/L MIC); $72.7 \text{ vs } 9.1\%, p = 0.01$ (total LNZ and 4 mg/L MIC); $100 \text{ vs } 36.4\%, p = 0.01$ (LNZu and 1 mg/L MIC); $90.9 \text{ vs } 27.3\%, p = 0.01$ (LNZu and 2 mg/L MIC), Fig. 2.

Comparing the two groups, patients receiving CI showed a trend toward higher mean ± SD $AUC_{0–24}$ and $AUC_{0–24,u}$ (146.3 ± 51.5 vs 110.6 ± 55.3 mg h/L, $p = 0.13$ and 101 ± 35.5 vs 76.34 ± 38.1, $p = 0.13$). Although not statistically significant, the percentage of patients with an $AUC_{0–24}/MIC$ (2 mg/L) ratio ≥80 was higher in the CI group (36.3 vs 18.2 %, $p = 0.64$), Table 2. For patients receiving II, $V_d$, $CL$, and $t_{1/2}$ were $45.1 \pm 18$ L, $14.3 \pm 7$ L/h, and $2.4 \pm 1$ h, respectively. In the two patients undergoing CRRT during CI, $CL_{CVVHDF}$, $C_{ss}$, and $AUC_{0–24}$ were $2.4/0.67$ L/h, $4.1/5.6$ mg/L, and $101.1/134.9$ h, respectively. The exclusion of these subjects from the CI group did not significantly change the PK results (see Table 1 in the ESM). Creatinine clearance in the remaining 20 patients was between 40 and 80 mL/min in two subjects (one for each group) and more than 120 mL/min in 13 subjects (5 in the CI group and 8 in the II group).

**ELF penetration**

Fourteen out of 22 patients underwent LNZ ELF concentrations determination: 7 in the CI group and 7 in the II group. LNZ did diffuse well into the lungs and the ELF/plasma penetration ratio (%) was slightly higher in the CI group [106 (IQR 71.6–116) vs 80 (IQR 56.6–130.5); $p = 0.46$].

However, using a Monte Carlo simulation, a significant difference was observed in the ELF/plasma penetration ratio (%) between the two groups [CI group, 98.8 (IQR 93.8–104.3) vs II group, 87.1 (IQR 78.7–95.4); $p < 0.001$] (Fig. 3).

**Discussion**

In critically ill obese patients with VAP, LNZ CI was more effective than II in obtaining PD parameters that predict its in vivo activity, even though for $AUC/MIC$ the difference did not reach statistically significant power. Intrapulmonary drug penetration was optimal in both groups, but CI appeared to have a better distribution in the lung.

Critically illness status may strongly influence the PK profile of many antibiotics. Variations of extracellular fluids and renal clearance are the main determinants of antimicrobial drug distribution and elimination [14]. In our II group, $C_{\text{max}}$ and $C_{\text{min}}$ were remarkably low, and the $AUC_{0–24}$ and $T > MIC$ values were inadequate to optimally treat MRSA strains with high MICs (2–4 mg/L). Our results are consistent with a randomized controlled trial which compared LNZ CI vs II in 16 septic patients [18], wherein Adembri et al. observed that in all the subjects receiving standard intermittent dosing the
Table 2 Steady-state serum and alveolar LNZ PK/PD parameters in the 22 enrolled patients

<table>
<thead>
<tr>
<th>Parameter</th>
<th>II group (n = 11)</th>
<th>CI group (n = 11)</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vd, L</td>
<td>45.1 ± 18.2</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>CL, L/h</td>
<td>14.3 ± 6.8</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>t1/2, h</td>
<td>2.4 ± 0.8</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Cmax mg/L</td>
<td>10 ± 3.7</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Cmax ss, mg/L</td>
<td>6.8 ± 2.6</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Cmin mg/L</td>
<td>1.7 ± 1.1</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Cmin ss, mg/L</td>
<td>1.2 ± 0.8</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>.Css mg/L</td>
<td>–</td>
<td>6.2 ± 2.3</td>
<td>–</td>
</tr>
<tr>
<td>Cmax ss, mg/L</td>
<td>–</td>
<td>4.3 ± 1.6</td>
<td>–</td>
</tr>
<tr>
<td>ELF/plasma ratio (%), median (IQR)</td>
<td>80 (56.6–130.5)</td>
<td>106 (71.6–116)</td>
<td>0.46</td>
</tr>
<tr>
<td>Cmax ss, median (IQR)</td>
<td>8.3 (6.7–9.8)</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Cmax ss, median (IQR)</td>
<td>–</td>
<td>5.3 (3.8–7.6)</td>
<td>–</td>
</tr>
<tr>
<td>AUC0-24, mg h/L</td>
<td>110.6 ± 55.3</td>
<td>146.3 ± 51.5</td>
<td>0.13</td>
</tr>
<tr>
<td>AUC0-24, mg h/L</td>
<td>76.3 ± 38.1</td>
<td>101 ± 35.5</td>
<td>0.13</td>
</tr>
<tr>
<td>AUC0-24, mg h/L</td>
<td>55.3 ± 27.6</td>
<td>73.2 ± 25.7</td>
<td>0.13</td>
</tr>
<tr>
<td>AUC0-24, mg h/L</td>
<td>38.2 ± 19.1</td>
<td>50.5 ± 17.8</td>
<td>0.13</td>
</tr>
<tr>
<td>AUC0-24, mg h/L</td>
<td>18.2</td>
<td>36.3</td>
<td>0.64</td>
</tr>
<tr>
<td>AUC0-24, mg h/L</td>
<td>–</td>
<td>0</td>
<td>–</td>
</tr>
<tr>
<td>% T &gt; 0.5 mg/L MIC, median (IQR)</td>
<td>100 (97.1–100)</td>
<td>100 (100–100)</td>
<td>0.23</td>
</tr>
<tr>
<td>% T &gt; 0.5 mg/L MIC, median (IQR)</td>
<td>100 (96.8–100)</td>
<td>100 (100–100)</td>
<td>0.22</td>
</tr>
<tr>
<td>% T &gt; 1 mg/L MIC, median (IQR)</td>
<td>100 (89–100)</td>
<td>100 (100–100)</td>
<td>0.05*</td>
</tr>
<tr>
<td>% T &gt; 1 mg/L MIC, median (IQR)</td>
<td>96.7 (68–100)</td>
<td>100 (100–100)</td>
<td>0.003*</td>
</tr>
<tr>
<td>% T &gt; 2 mg/L MIC, median (IQR)</td>
<td>100 (100–100)</td>
<td>100 (100–100)</td>
<td>0.009*</td>
</tr>
<tr>
<td>% T &gt; 2 mg/L MIC, median (IQR)</td>
<td>66.3 (39.3–95.8)</td>
<td>100 (100–100)</td>
<td>0.006*</td>
</tr>
<tr>
<td>% T &gt; 4 mg/L MIC, median (IQR)</td>
<td>100 (74.2–100)</td>
<td>100 (74.2–100)</td>
<td>0.005*</td>
</tr>
<tr>
<td>% T &gt; 4 mg/L MIC, median (IQR)</td>
<td>21.2 (16.3–55.3)</td>
<td>0 (0–100)</td>
<td>0.72</td>
</tr>
</tbody>
</table>

Data are expressed as mean ± SD unless otherwise indicated. LNZ linezolid, PK/PD pharmacokinetic/pharmacodynamic, II intermittent infusion, CI continuous infusion, Vd volume of drug distribution, IQR interquartile range, CL drug clearance, t1/2 elimination half-life, Cmax peak plasma concentration, Cmin trough plasma concentration, Cmax ss unbound peak plasma concentration, Cmin ss unbound trough plasma concentration, Css steady-state plasma concentration, ELF epithelial lining fluid, MIC minimum inhibitory concentration, AUC/AUCss total drug/unbound area under the time-concentration curve, T > MIC time above the minimum inhibitory concentration, Tss > MIC time above the minimum inhibitory concentration (unbound fraction), – not applicable, BAL bronchoalveolar lavage
* p ≤ 0.05
† BALs were collected in 7 patients 2 h after of the fifth infusion (peak concentration on day 3 of treatment)
‡ BALs were collected in 7 patients on day 3 of treatment at the mid-interval (53 or 57 h)

mean trough levels (both total and free) were below 4 mg/mL and in half of them less than 1 mg/mL. On the other hand all the patients in the CI group showed unbound and total LNZ Css above the susceptibility threshold. In our patients undergoing CI, although not significantly, mean AUC0-24 values for both total and unbound LNZ were higher than in the II group, showing values about 100 mg h/L. In the same way, CI provided significantly higher T > MIC and PTA than II, but these advantages were not so evident for the extreme values of the MIC susceptibility range (0.5 mg/L and 4 mg/L).

In our study LNZ t1/2 was lower than previously reported in healthy subjects (2.4 ± 0.8 h). However many pathophysiological changes (i.e., increased cardiac output, leaky capillaries, augmented renal clearance, low protein concentration and altered bounding) occurring in severe critically ill patients may modify antibiotics’ PKs, increasing their body clearance. Indeed, septic patients studied by Adembri et al. showed similar t1/2 values (3.5 ± 2.2 h).

Continuous infusion is a simple strategy to optimize the duration of exposure above the MIC for time-dependent antibiotics. Concentrations up to 4–5 times the MIC, increasing the AUC value, may maximize killing activity, but higher values do not add any benefits [30]. Few data are available on LNZ CI use. In addition to the study by Adembri et al. the only other report addressing this issue was recently published by Boselli et al. [13]. These authors, in a cohort of 12 ICU patients undergoing LNZ CI, describedCss, AUC0-24 and alveolar penetration values similar to those we observed in our cohort [7.1 mg/L (6.1–9.8), 169 mg h/L (146–235), 97 % (80–108), respectively] [13]. However in this study neither obese nor hyperfiltrating patients were included and no detail about their severity degree was provided (i.e., SOFA score, presence of septic shock).

Our critically ill patients were moderately obese [median BMI (IQR) 33.2 kg/m² (32.6–37.5)]. Obesity may significantly influence antibiotics’ PK, but few clinical data are available in this field. Vd is modified by...
the increase in both lean body weight and adipose tissue. Furthermore kidney mass and the correspondent global filtration may influence drugs’ CL. Linezolid is a moderately lipophilic drug and, in our patients, both critically ill status and obesity could have impaired the PK profile during II administration. This detrimental effect was blunted by the adoption of continuous infusion.

Subtherapeutic LNZ concentrations have been reported after bolus administration in obese patients. Both increased CL and $V_d$ have been previously observed [23, 31]. Different results were recently shown after orally intermittent LNZ administration to 20 healthy obese (moderately and morbidly) volunteers [22]. In that paper mean $\pm$ SD AUC$_{0-12}$ and $C_{\text{max}}$ values (119.8 $\pm$ 46.24 mg h/L and 19.8 $\pm$ 4 mg/L, respectively) were adequate to ensure optimal bacterial killing and mean $\pm$ SD $V_d$ value (44.1 $\pm$ 9.9 L) was comparable to normal weight subjects. In any case, a significant positive relationship between the body weight and AUC values was found ($r^2$ $< 0.5$). Our data are partially in accordance with what was stated previously. After II, we observed lower $C_{\text{max}}$, $C_{\text{min}}$, and AUC$_{0-24}$ than previously reported but the $V_d$ was not so increased (45.1 $\pm$ 18.2 L). This finding may be explained by the low obesity degree of our patients whose total body weight was less than 150 kg [32]. On the other hand our patients showed high creatinine clearance values which certainly have contributed to the low observed LNZ concentrations in the II group. However calculated creatinine clearance may be not appropriate to identify augmented renal clearance in septic patients. Additionally our results may not be applied to morbidly obese patients where a larger $V_d$ is supposed to further modify LNZ PK.

Our report is the first to investigate LNZ pulmonary distribution in obese critically ill patients according to infusion modality. It is well known that LNZ optimally penetrates the lung and this PK property has been recently confirmed in 12 critically ill patients receiving CI [13]. Our study confirms this PK property, in a population of moderately obese critically ill patients receiving either II or CI infusion. However, after performing a Monte Carlo simulation, CI was associated with a higher median ELF/plasma ratio percentage [CI group, 98.8 (IQR 93.8–104.3) vs II group, 87.1 (IQR 78.7–95.4); $p < 0.001$], resulting in ELF $C_{\text{ss}}$ above 4 mg/L in all studied patients. However, the limited number of analyzed samples (7 for each group) and the absence of AUC$_{0-24}$ data (every patient underwent a single BAL sampling) certainly reduce the significance of the difference showed by our simulation.

Mean SAPS II values were not similar between the two groups. The presence of few outliers in a small sample sized PK study is the main reason for this heterogeneity. However, the most relevant clinical variables that correlated with our endpoint (BMI, septic shock, albumin concentration, renal function) were homogeneously distributed.

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**Fig. 2** Probability of target attainment of pharmacodynamic indices (a 85% $T > \text{MIC}$, b 100% $T > \text{MIC}$) in plasma for intermittent infusion and continuous infusion (unbound and total drug). CI continuous infusion total drug, CI$_u$ continuous infusion unbound drug, II intermittent infusion total drug, II$_u$ intermittent infusion unbound drug, MIC minimum inhibitory concentration. Samples were collected on day 3 of treatment.

**Fig. 3** Box plot showing percentage differences between II and CI LNZ ELF/plasma ratio. The results are based on a Monte Carlo simulation with 1,000 iterations. Boxes represent interquartile ranges (lower border 25th percentile; upper border 75th percentile), and the horizontal lines within the boxes indicate the medians (50th percentile). Whiskers indicate minimum and maximum values. CI continuous infusion total drug, II intermittent infusion total drug, ELF epithelial lining fluid. Samples were collected on day 3 of treatment.
Finally we did not identify any LNZ-related AE. This is not surprising, since the observed concentrations were far from the safety thresholds ($C_{\text{min}}$ 10 mg/L and AUC$_{0-24}$ 400 mg h/L) [10, 11]. In addition we excluded patients who were receiving drugs which could interfere with LNZ metabolism.

This study has some limitations. First, our population was represented by moderately obese patients and the results may not be generalized to subjects with BMI higher than 40 kg/m$^2$. Secondly, we could perform only 14 out of the 22 planned BALs. Thirdly, the number of Gram-positive VAP, the duration of CI administration, and some baseline differences (i.e., SAPS II, PaO$_2$/FiO$_2$ ratio) do not allow us to address any conclusive clinical consideration.

However, to the best of our knowledge, this is the first randomized trial investigating the plasma and pulmonary PK profile of LNZ CI administration, compared to II, in critically ill obese patients with VAP.

Conclusions

In summary, II LNZ administration in obese critically ill patients with VAP is associated with suboptimal plasma concentrations. CI administration is able to safely improve the LNZ PK profile but it may still be inadequate for the management of difficult to treat germs (i.e., MRSA with a MIC of 4 mg/L). Critically ill status and obesity do not strongly affect pulmonary distribution but CI provides a higher alveolar penetration ratio. Clinical trials are needed to verify the potential clinical advantages of LNZ CI in ICU patients at risk of antibiotic underexposure.

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References


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